

# Mouse IL-10 Precoated ELISA Kit

Catalog Number: EXMK008

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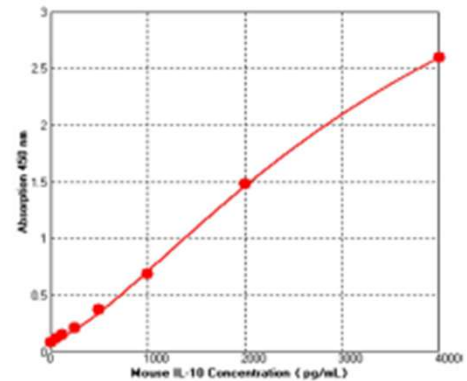
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For Research Use Only. Not Intended for Diagnostic or Therapeutic Use.

## Product Description

The Mouse IL-10 Precoated ELISA Kit is used for the *in vitro* quantitative detection of mouse IL-10 in mouse serum, plasma, buffer or cell culture medium. Natural and recombinant mouse IL-10 are captured by the immobilized antibody coated on the microplate and detected by the biotinylated IL-10 antibody followed by streptavidin-HRP and the tetramethylbenzidine (TMB) substrate. Read this manual in its entirety before use.

- Assay time: 2 hours 20 minutes
- Detection range: 62.5-4000 pg/mL
- Sensitivity: 32 pg/mL
- Precision: Intra-assay CV: ≤10%; Inter-assay CV: ≤15%
- Specificity: no cross-reactivity with mouse cytokines



Example Standard Curve

## Limitations

- For Research Use Only. Not Intended for Diagnostic or Therapeutic Use.
- Variations in temperature, incubation time or technique while performing the assay can affect results.
- Variations in collecting, processing and storing samples may affect sample values.
- Do not substitute kit components with reagents from other sources or lots.

## Materials Provided, Storage and Reconstitution

Store the kit at 2-8°C for up to 12 months. Do not use past kit expiration date.

Reagents	Quantity	Reconstitution
<b>Mouse IL-10 Standard, lyophilized</b>	2 vials	Follow instructions on the vial. <b>Use a new standard for each assay. Discard after use.</b>
<b>Biotinylated Mouse IL-10 Antibody (25 µL)</b>	2 vials	1:100 dilution with Dilution Buffer R (1X)
<b>Streptavidin-HRP (50 µL)</b>	2 vials	1:100 dilution with Dilution Buffer R (1X)
<b>Dilution Buffer R (1X) (10mL)</b>	3 bottles	Ready-to-use
<b>Washing Buffer (50X) (15 mL)</b>	1 bottle	1:50 dilution with distilled water
<b>Color Reagent (TMB) (10 mL)</b>	1 bottle	Ready-to-use
<b>Stop Solution (10 mL)</b>	1 bottle	Ready-to-use
<b>Precoated Mouse IL-10 96-well ELISA plate</b>	1 plate (12 strips of 8 wells)	Ready-to-use
<b>Plate Sealers</b>	4 strips	Ready-to-use

## Additional Required Supplies

- Microplate reader with 450 nm filter (with optional wavelength correction filter at 620 nm)
- Adjustable pipettes and pipette tips
- Distilled or deionized water
- Vortex mixer and magnetic stirrer.

## Precautions

- Store the kit according to the labels and instructions.
- Reagents should be at room temperature before use.
- Streptavidin-HRP and biotinylated antibody should be centrifuged briefly before use.
- Store unused plate strips in the sealed pouch.
- Avoid cross-contamination: change pipette tips between adding each standard dilution, between samples, and between reagents.
- Use separate reservoirs for individual reagents.
- Ensure reagents are mixed evenly by shaking or stirring. Avoid foaming of reagents.
- Add all reagents in the same order to the plate to ensure consistent reaction times for each step.
- Remove remaining wash buffer from wells completely after each wash step. Blot the inverted plate on clean filter paper or paper towels before adding the next reagent.
- Color Reagent (TMB) is sensitive to light and exposure to metals. Protect from light and avoid contact with metals. If unused Color Reagent is contaminated, it will turn blue before use and should be discarded appropriately.
- For each experiment, a standard curve must be generated.
- Results outside the linear range of the standard curve are not accurate. Dilute samples generating values above the highest standard with Dilution Buffer R (1X).

## Preparation of Reagents

**Mouse IL-10 Standard:** Please refer to the product label for reconstitution instructions. Let stock solution in reconstituted vial stand for 5 minutes. Shake gently until completely dissolved.

**Dilution Step (refer to table below):** Label 8 test tubes S0-S7. Once dissolved, transfer the Mouse IL-10 Standard stock solution to the S7 tube. Add 250  $\mu$ L of Dilution Buffer R (1X) to tubes S0-S6. Pipette 250 $\mu$ L of the standard stock solution from tube S7 into tube S6 for a 2-fold dilution. Mix each dilution thoroughly and continue. Perform a 2-fold serial dilution for tubes S5-S1 by pipetting 250 $\mu$ L into each successive tube. Tube S0 serve as the blank control and should only contain Dilution Buffer R (1X).

Standard Tube Number	Dilution Ratio	Dilution Buffer R (1X) ( $\mu$ L)	Standard Concentration (pg/mL)
S7	---	---	4000
S6	1:2	250	2000
S5	1:4	250	1000
S4	1:8	250	500
S3	1:16	250	250
S2	1:32	250	125
S1	1:64	250	62.5
S0	---	250	0

**Biotinylated Mouse IL-10 Antibody:** Dilute 1:100 with Dilution Buffer R (1X) in a clean tube according to the table below to prepare the 1X working solution. Mix well.

Number of Wells	Biotinylated Antibody ( $\mu$ L)	Dilution Buffer R (1X) ( $\mu$ L)
16	9	900
24	13	1300
32	17	1700
48	25	2500
96	50	5000

## Preparation of Reagents (continued)

**Streptavidin-HRP:** Dilute 1:100 with Dilution Buffer R (1X) in a clean tube according to the table below to prepare the 1X working solution. Mix well.

Number of Wells	Streptavidin-HRP ( $\mu\text{L}$ )	Dilution Buffer R (1X) ( $\mu\text{L}$ )
16	17	1700
24	25	2500
32	34	3400
48	50	5000
96	100	10000

**Wash Buffer (50X):** Dilute 1:50 with distilled water to prepare 1X solution.

## Protocol

1. Reagents and samples should be at room temperature before use.
2. Determine the required number of wells required for the samples, standard curve, and blank control to be tested in duplicate. Remove unneeded plate strips and store in sealed pouch.
3. Add 100  $\mu\text{L}$  of standard and sample per well.
4. Add 100  $\mu\text{L}$  of Dilution Buffer R (1X) from tube S0 to the blank control wells.
5. Add 50  $\mu\text{L}$  of Biotinylated Mouse IL-10 Antibody (1X) to each well. Cover with the plate sealer and incubate for 90 minutes at room temperature (18-25°C).
6. Aspirate each well. Add 300  $\mu\text{L}$  of Wash Buffer (1X) to each well. Let stand for 1 minute. Repeat 3 times. Remove remaining wash buffer from wells completely after each wash step. Blot the inverted plate on clean filter paper or paper towels before add the next reagent.
7. Add 100  $\mu\text{L}$  of streptavidin-HRP (1X) to each well. Cover with the plate sealer and incubate for 30 minutes at room temperature.
8. Repeat the wash steps, see Step 6.
9. Add 100  $\mu\text{L}$  of Color Reagent to each well. Incubate for 10-20 minutes at room temperature. **Protect from light.**
10. Add 100  $\mu\text{L}$  of Stop Solution to each well. A color change from blue to yellow should be observed. Gently tap the side of the plate to ensure thorough mixing.
11. Within 10 minutes, read the absorbance values using a microplate reader at 450 nm, with wavelength correction at 620 nm. The wavelength correction compensates for optical imperfections of the plate. Values measured at 450 nm without correction will be less accurate.

## Data Analysis

Calculate the average optical density (O.D.) of the duplicate wells for the standards, samples and controls. Subtract the average blank control O.D value from all wells.

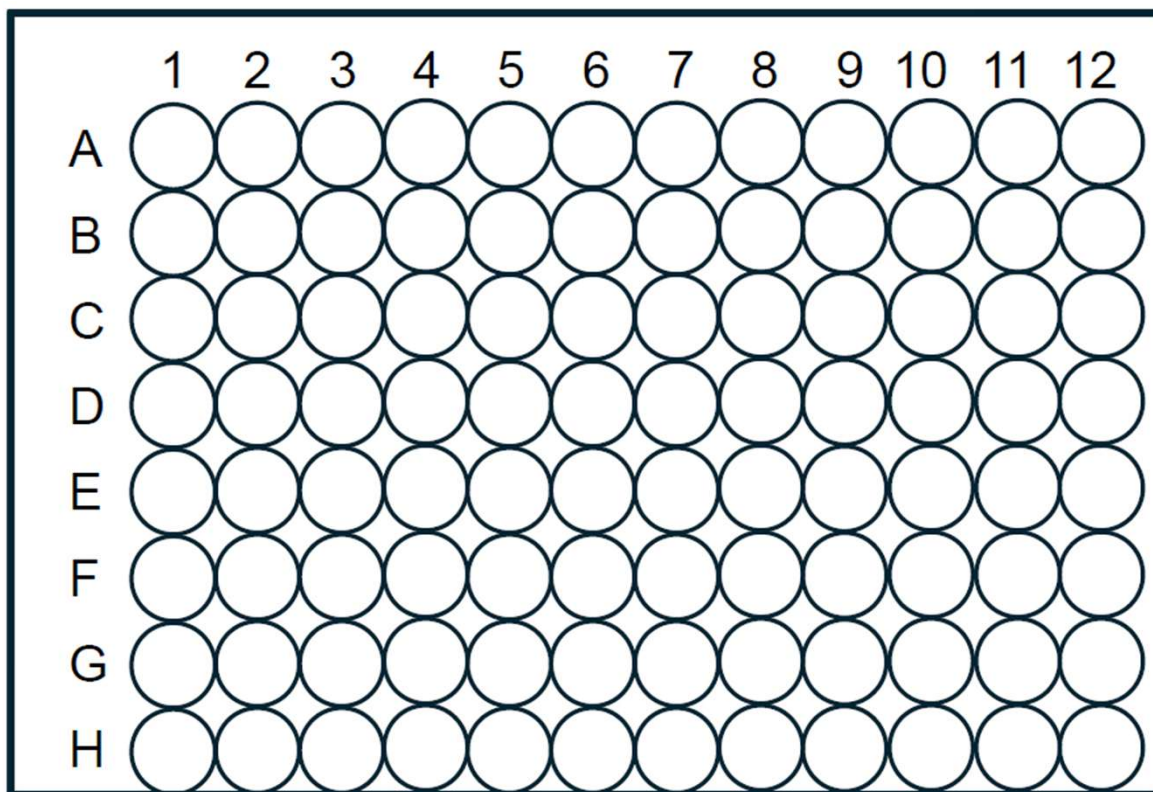
With suitable software, generate the standard curve using a four parameter (4-PL) curve-fit. The standard concentration is plotted on the x-axis and the mean O.D. value is plotted on the y-axis.

The software will report the calculated concentrations of the sample wells based on the standard curve. If the samples were diluted prior to adding to the wells, these calculated concentrations must be multiplied by the dilution factor.

Problem	Possible Source	Suggested Action
Weak signal across entire plate	Incorrect washing method	Verify correct washing method and wash buffer
	Reagents have expired	Repeat assay with fresh reagents
	Streptavidin-HRP concentration is too low	Repeat assay according to recommended dilution of streptavidin-HRP
	Insufficient incubation time	Repeat assay according to recommended incubation time of each step
	Incorrect storage of reagents	Store reagents per recommended temperature, time and protect from light, as noted.
	Incorrect wavelength in microplate reader	Check filters in microplate reader
Poor duplicates	Incorrect washing method	Verify correct washing method and wash buffer
	Plates are not clean	Wipe the bottom of the plate before reading the plate
	Incorrect wavelength in microplate reader	Check filters in microplate reader
	Reagents from different lots or sources were substituted in the kit	Only use reagents supplied in the kit
	Cross-contamination in the well	Use new pipette tips between samples, standards, and reagents
	Plate sealer reused or omitted	Use fresh plate sealer for each incubation
No signal (positive control or standard curve does not develop color)	Contamination of streptavidin-HRP with sodium azide	Use fresh reagents
	Protocol steps omitted or incorrect order	Review protocol and repeat with correct addition of reagents
	Reagents have expired	Repeat assay with fresh reagents
High background	Incorrect washing method	Verify correct washing method and wash buffer
	Color Reagent is contaminated or exposed to light	Verify Color Reagent is colorless before use and is stored properly
	Reagents have expired	Repeat assay with fresh reagents
	Incorrect reagent dilution, e.g. streptavidin-HRP or biotinylated antibody concentration too high	Repeat assay according to recommended dilution
	Contamination of wells with standards or positive samples	Repeat assay and avoid cross-contamination of wells
	Incubation time too long or temperature is too high	Repeat assay according recommended incubation time and temperature

## Plate Layout

This template is provided to record layout of standards and samples.



## Protocol At-A-Glance

Add 100  $\mu$ L sample, standard and blank (diluent only) to each well



Add 50  $\mu$ L diluted biotinylated antibody to each well



Incubate for 90 minutes at room temperature and wash 3 times



Add 100  $\mu$ L diluted streptavidin-HRP to each well



Incubate for 30 minutes at room temperature and wash 3 times



Add 100  $\mu$ L of ready-to-use TMB to each well and incubate for 10-20 minutes (protect from light)



Add 100  $\mu$ L of Stop Solution to each well



Read the plate at 450 nm (620 nm correction) within 10 minutes